

Department of Biochemistry, UCS, Osmania University

Approved MSc. Biochemistry CBCS Syllabus

(Effective from 2022-23 admitted batch)

SEMESTER-I

PAPER	TITLE	Credits*	hrs/ week	Internal marks( Theory)	Final exam marks(T heory)	Practical exam marks
I	<b>BI101:</b> Chemistry of Biomolecules (core)	5 (3T+2P)	7	30	70	50
II	<b>BI102:</b> Endocrine Biochemistry, Vitamins and Nucleic acids (core)	5 (3T+2P)	7	30	70	50
III	<b>BI103:</b> Cell Biology and Bioenergetics (core)	5 (3T+2P)	7	30	70	50
IV	<b>BI 104:</b> Basic Bioanalytical Techniques (core)	5 (3T+2P)	7	30	70	50
<b>TOTAL</b>		<b>20</b>	<b>28</b>	<b>400</b>		<b>200</b>

\*One credit means the standard methodology of calculating **one hour** of theory or one hour of tutorial or **two hours of laboratory work**, per week for a duration of a semester (13-15 weeks)

**PAPER-I BI 101: CHEMISTRY OF BIOMOLECULES**

**(5 credits: theory credits= 3+ practical credits= 2)**

<b>THEORY</b>	
<b>CREDIT 1</b>	<b>Amino Acids, &amp; Proteins (15h)</b>
	<ol style="list-style-type: none"><li>1. Classification and structure of amino acids,</li><li>2. Essential, non essential, and non protein or unusual amino acid.</li><li>3. General properties and Acid- Base Reactions of AA, (pKa Values)</li><li>4. Peptide bond – stability and formation, polypeptides.</li><li>5. Methods for determining amino and carboxy terminal and molecular weight</li><li>6. Primary structure of proteins</li><li>7. Secondary structure <math>\alpha</math> helix, <math>\beta</math> sheet, 310 helix</li><li>8. GN Ramachandran plots: <i>Phi</i>, <i>Psi</i> and <i>omega</i> angle</li><li>9. Tertiary &amp; Quaternary structure (myoglobin, hemoglobin)</li><li>10. Small peptides (glutathione, peptide hormones)</li><li>11. Cyclic peptides (Gramicidin)</li><li>12. Classification of proteins - globular, fibrous</li><li>13. Membrane, Metallo - proteins, SCOP, CATH</li><li>14. Denaturation (pH, temperature, chaotropic agents), renaturation</li><li>15. Protein folding, role of chaperones in folding</li></ol>
<b>CREDIT 2</b>	<b>Carbohydrates (15h)</b>
	<ol style="list-style-type: none"><li>1. Classifications and structure of Carbohydrates</li><li>2. Configurations and conformations</li><li>3. Reactions of Monosaccharides</li><li>4. Stability and formations of glycosidic bond</li><li>5. Disaccharides and Oligosaccharides</li><li>6. Polysaccharides Structural (Cellulose, Chitin)</li><li>7. Storage (Starch, Glycogen, Inulin)</li><li>8. Hemicelluloses- Lignin, Pectin</li><li>9. Hetero-Polysaccharides/ acidic Mucopolysaccharides Glycosaminoglycan</li><li>10. Chemistry and biological role of Hyaluronic acids, Chondroitin sulphate,</li><li>11. Keratansulphate, dermatansulphate, heparin</li><li>12. Glycoproteins and Proteoglycans</li><li>13. Bacterial cell wall Polysaccharides, Peptidoglycans</li><li>14. Blood group glycoproteins</li><li>15. Structural determination of polysaccharides</li></ol>
<b>CREDIT 3</b>	<b>Lipids &amp; Porphyrins (15h)</b>
	<ol style="list-style-type: none"><li>1. Classification of lipids &amp; fatty acids</li><li>2. Biological significance of lipids &amp; fatty acids</li><li>3. Steroids, Sterols, relation to vitamin D</li><li>4. Steroid hormones</li><li>5. Bile acids and salts</li><li>6. Phospholipids</li><li>7. Oils, waxes, isoprene units</li><li>8. Lipoproteins</li><li>9. Glycolipids</li><li>10. Sphingolipids</li><li>11. Structure &amp; function of porphyrins : Heme</li><li>12. Structure &amp; function of porphyrins : Chlorophyll</li><li>13. Cerebrosides, Gangliosides</li><li>14. Prostaglandins, Prostacyclins, Eicosanoids</li><li>15. Thromboxanes, Leukotrienes</li></ol>

<b>PRACTICAL (Each practical topic consists of 3 Experiments, Total 15 Expts)</b>	
<b>CREDIT 4</b>	<b>Amino acid analysis (30 h)</b>
	<ol style="list-style-type: none"> <li>1. Qualitative analysis of amino acids</li> <li>2. Determine pKa and pI of acidic, basic, and neutral amino acids</li> <li>3. Estimation of amino acids by Ninhydrin methods</li> <li>4. Quantification of glycine by formal titration</li> <li>5. Estimation of tryptophan by Spies and Chambers method</li> </ol>
<b>CREDIT 5</b>	<b>Lipid analysis (30 h)</b>
	<ol style="list-style-type: none"> <li>1. Qualitative analysis of lipids</li> <li>2. Saponification value of fats</li> <li>3. Iodine number of oil</li> <li>4. Peroxide value of fats</li> <li>5. Acid value of fats</li> </ol>

**PAPER-II BI 102 : ENDOCRINE BIOCHEMISTRY, VITAMINS AND NUCLEIC ACIDS**

**(5 credits: theory credits= 3+ practical credits= 2)**

<b>THEORY</b>	
<b>CREDIT 1</b>	<b>Endocrine System (15h)</b> <ol style="list-style-type: none"><li>1. Endocrine glands and their hormones</li><li>2. Control of hormone secretion</li><li>3. Mechanism of hormone action</li><li>4. Pituitary gland: Structure, Anterior Pituitary : hormones and their functions</li><li>5. Posterior Pituitary- Hormones and their functions</li><li>6. Disorders related to pituitary hormones</li><li>7. Thyroid gland and Parathyroid gland: Structure and functions</li><li>8. Disorders: hypothyroidism, hyperthyroidism</li><li>9. Parathormone and associated disorders</li><li>10. Adrenal gland: Structure</li><li>11. Secretions of adrenal cortex and their functions, hypoadrenalism, hyperadrenalism</li><li>12. Secretions of adrenal medulla and their functions</li><li>13. Pancreas: Islets of Langerhans, alpha and beta cells</li><li>14. Functions of Insulin and glucagon, deficiency of insulin</li><li>15. Testes and Ovaries Structure, functions of testosterone, estrogens and progesterone</li></ol>
<b>CREDIT 2</b>	<b>Vitamins (15h)</b> <ol style="list-style-type: none"><li>1. Water Soluble Vitamins: Structure and Classification</li><li>2. Water Soluble Vitamins: Chemistry, Biological Source and physiological significance</li><li>3. Fat Soluble Vitamins: Structure and Classification</li><li>4. Fat Soluble Vitamins: Chemistry, Biological Source and Significance</li><li>5. Structure, function and the deficiency disorders of Vit B1 (Thiamine), B2 (Riboflavin)</li><li>6. Structure, function and the deficiency disorders of and B3 (Niacin) and Vit B5 (Pantothenic acid)</li><li>7. Structure, function and the deficiency disorders of B6 (Pyridoxine) and B7 (Biotin)</li><li>8. Structure, function and the deficiency disorders of Vit B9 (Folic acid) and B12 (Cobalmins)</li><li>9. Structure, function and the deficiency disorders of Vit C (Ascorbic acid)</li><li>10. Structure, function and the deficiency disorders of Vit A (Retinol)</li><li>11. Structure, function and the deficiency disorders of Vit D (Calciferol)</li><li>12. Structure, function and the deficiency disorders of Vit E (Tocopherol)</li><li>13. Structure, function and the deficiency disorders of Vit K (Phytonadione)</li><li>14. Recommended daily allowance of vitamins</li><li>15. Vitamin supplementation</li></ol>
<b>CREDIT 3</b>	<b>Chemistry of Nucleic acids (15h)</b> <ol style="list-style-type: none"><li>1. Purines: structure and function</li><li>2. Pyrimidines: structure and function</li><li>3. Nucleosides, nucleotides, phospho-diester bond</li><li>4. Unusual bases, Modified bases: structure and properties</li><li>5. Structure of DNA – Watson Crick Model A – and Z - forms</li><li>6. Supercoiling of DNA – negative and positive, linking number</li><li>7. Properties of DNA denaturation and renaturation</li><li>8. <math>T_m</math> ( factors affecting <math>T_m</math> ) and <math>C_{0t}</math> curves</li><li>9. Structure of RNA: mRNA, tRNA</li><li>10. Structure of RNA: rRNA, siRNA and miRNA</li><li>11. Properties of RNA denaturation and renaturation</li><li>12. Difference between DNA and RNA</li><li>13. Heteroduplex mapping</li><li>14. D loops and R loops</li><li>15. Catalytic RNA</li></ol>

<b>PRACTICAL (Each practical topic consists of 3 Experiments, Total 15 Expts)</b>	
<b>CREDIT 4</b>	<b>Carbohydrate analysis (30 h)</b>
	<ol style="list-style-type: none"> <li>1. Qualitative analysis of carbohydrates</li> <li>2. Quantitative analysis of carbohydrates</li> <li>3. Estimation of Fructose</li> <li>4. Estimation of total sugars by phenol sulfuric acid method</li> <li>5. Estimation of reducing sugars by DNS</li> </ol>
<b>CREDIT 5</b>	<b>Nucleic acid analysis (30 h)</b>
	<ol style="list-style-type: none"> <li>1. Estimation of DNA by DPA</li> <li>2. Assessment of DNA purity by A260/A280 method</li> <li>3. Estimation of RNA by Orcinol method</li> <li>4. Separation of purines by paper chromatography</li> <li>5. Separation of pyrimidines by paper chromatography</li> </ol>

**PAPER-III BI 103 : CELL BIOLOGY AND BIOENERGETICS**

**(5 credits: theory credits= 3+ practical credits= 2)**

<b>THEORY</b>	
<b>CREDIT 1</b>	<b>Structure of Prokaryotic &amp; Eukaryotic cells (15 h)</b>
	<ol style="list-style-type: none"><li>1. Classification of prokaryotes and eukaryotes (systems of classification)</li><li>2. Ultrastructure of eubacteria, cyanobacteria, mycoplasma</li><li>3. Ultra structure of plant cell</li><li>4. Ultrastructure of animal cell</li><li>5. Composition of cytoskeleton: microfilaments, microtubules, intermediate filaments</li><li>6. Function of cytoskeleton ( transport and cell division)</li><li>7. Nuclear skeleton : lamina scaffold</li><li>8. Structure of chromosome ( centromere, telomere, kinetochore )</li><li>9. Euchromatin &amp; heterochromatin</li><li>10. Formation and structure of special chromosomes ( polytene and lampbrush)</li><li>11. Cell cycle</li><li>12. Mitosis and Meiosis</li><li>13. Cell cycle check points and regulation</li><li>14. Apoptosis</li><li>15. Regulation of apoptosis</li></ol>
<b>CREDIT 2</b>	<b>Bio Membranes ( 15 h)</b>
	<ol style="list-style-type: none"><li>1. Composition and Structure of Cell membrane</li><li>2. Membrane Dynamics</li><li>3. Membrane Lipids: Composition distribution and functions</li><li>4. Membrane Proteins: Composition distribution and functions</li><li>5. Trans membrane proteins and their classification</li><li>6. Methods of detecting Trans membrane proteins, Hydropathy plots</li><li>7. Membrane Asymmetry</li><li>8. Fluid Mosaic Model of Membrane</li><li>9. Membrane fluidity and its regulations, Flip flop.</li><li>10. RBC membrane structure</li><li>11. Membrane transport: active and passive transport, symport and antiport; NaK ATPase</li><li>12. Transport channels: voltage gated, ion gated and ligand-gated channels</li><li>13. Aquaporin, Glucose transporters, valinomycin: structure and mechanism of action</li><li>14. Artificial membranes: Reconstitution of functional membrane system from purified components</li><li>15. Liposomes, Micelles and vesicles</li></ol>
<b>CREDIT 3</b>	<b>Bioenergetics (15 h)</b>
	<ol style="list-style-type: none"><li>1. Elements of importance in Biochemistry (H,C,N,O,P,S), types and energy of bonds and interactions (ionic, covalent, coordinate, H-bonds, van der Waals, hydrophobic interactions )</li><li>2. Laws of thermodynamics, Gibbs free energy, relevance of entropy and enthalpy in biological systems and reactions.</li><li>3. Thermodynamically coupled reactions</li><li>4. Order of the reactions: first and second order reactions</li><li>5. Log and Incalesin biological processes (exponential growth curves, radioactivedecay)</li><li>6. Biological oxidation, Redox potential, Nernst equation</li><li>7. Enzymes involved in biological oxidation</li><li>8. High energy compounds</li><li>9. Oxidative phosphorylation</li><li>10. High energy bonds, phosphate potential, Forces stabilizing membrane</li><li>11 ETC in mitochondria</li><li>12 ETC in chloroplasts,</li><li>13. Un-couplers and inhibitors of energy transfer.</li><li>14. Shuttle pathways: Glycerol phosphate shuttle, Malate-Aspartate shuttle</li><li>15. Biological fluorescence (GFP and derivatives ), Bioluminescence</li></ol>

<b>PRACTICAL (Each practical topic consists of 3 Experiments, Total 15 Expts)</b>	
<b>CREDIT 4</b>	<b>Laboratory maintenance, safety and basic methods (30 h)</b>
	<ol style="list-style-type: none"> <li>1. GLP: Use of balance and pH meter, Lab safety,</li> <li>2. Calculations and preparation of standard solutions (primary, working standard)</li> <li>3. Preparation of buffers: Phosphate, Citrate and Tris buffer</li> <li>4. Titration: Estimation of Calcium in milk</li> <li>5. Titration: Estimation of Vitamin C in lemon juice</li> </ol>
<b>CREDIT 5</b>	<b>Separation and identification of biomolecules (30 h)</b>
	<ol style="list-style-type: none"> <li>1. SDS PAGE for protein</li> <li>2. Agarose gel for DNA</li> <li>3. Desalting of proteins by dialysis</li> <li>4. Gel filtration (size exclusion)</li> <li>5. Cell fractionation (centrifuge)</li> </ol>

**PAPER-IV BI 104 : BASIC BIOANALYTICAL TECHNIQUES**

**(5 credits: theory credits= 3+ practical credits= 2)**

<b>THEORY</b>	
<b>CREDIT 1</b>	<b>Spectroscopy and Chromatography (15h)</b> <ol style="list-style-type: none"><li>1. Colorimetry, Beer Lambert's Law-application and limitation, Molar extinction coefficient, Absorption maximum,</li><li>2. UV - Vis: Spectrophotometer – Instrumentation, application</li><li>3. Fluorescence Spectroscopy – principle, instrumentation, application</li><li>4. Infra red(IR) spectroscopy - principle, instrumentation, application</li><li>5. Raman spectroscopy - principle, instrumentation, application</li><li>6. CD – and ORD – principle, instrumentation, application</li><li>7. Partitioning and counter current distribution;</li><li>8. PC and TLC –principle ,instrumentation ,application</li><li>9. GC – principle, instrumentation, application</li><li>10. Ion – exchange chromatography – principle, instrumentation, application</li><li>11. Gel filtration (Gel exclusion chromatography ) – principle, application</li><li>12. Affinity chromatography: principle, instrumentation, application; immunoprecipitation</li><li>13. HPLC and RP–HPLC – principle, instrumentation, application</li><li>14. FPLC, LC – principle, instrumentation, application</li><li>15. Peptide mapping and N – terminal sequencing of proteins</li></ol>
<b>CREDIT 2</b>	<b>Centrifugation, Electrophoresis and Tracer techniques (15h)</b> <ol style="list-style-type: none"><li>1. Centrifugation, RCF and types of rotors Ultra centrifugation – principle, instrumentation,application</li><li>2. CsCl density gradient and sucrose gradient centrifugation – principle, application</li><li>3. Electrophoresis – moving boundary and zonal electrophoresis, Native and SDS PAGE</li><li>4. IEF and 2D PAGE,PAGE for DNA sequencing</li><li>5. AgaroseGels, PFGE Zymography</li><li>6. Denaturing gels for RNA, Southern and Northern Blots</li><li>7. Western Blot</li><li>8. Stable and radioactive isotopes, theory of radioactivity</li><li>9. Half life and emission spectra of biologically useful isotopes:<sup>2</sup>H, <sup>3</sup>H, <sup>14</sup>C, <sup>18</sup>O, <sup>32</sup>P, <sup>35</sup>S, <sup>125</sup>I; Isotopes used for labelling proteins (<sup>3</sup>H <sup>14</sup>C, <sup>35</sup>S, <sup>125</sup>I) and nucleic acids (<sup>3</sup>H, <sup>32</sup>P)</li><li>10. Detection of radioactivity by Scintillation counting Autoradiography</li><li>11. GM counter, gamma counter</li><li>12. Fluorography, applications of Phosphorimaging, luxometry and chemiluminescence as alternative to radioactivity</li><li>13. Radiation hazards and safe disposal of radio activity waste</li><li>14. Isotope dilution method – pulsechase</li><li>15. Historic examples -<sup>14</sup>C and <sup>18</sup>O to study photosynthesis; <sup>32</sup>P and <sup>35</sup>S to study viral replication ( Hershey – Chase experiment ), <sup>14</sup>N and <sup>15</sup>N in DNA replication (Meselson and Stahl experiment)</li></ol>
<b>CREDIT 3</b>	<b>Microscopy and Methods of Cell Study (15h)</b> <ol style="list-style-type: none"><li>1. Light Microscopy: Simple and compound microscope</li><li>2. Phase contrast microscopy</li><li>3. Dark field and polarization microscopy</li><li>4. Electron microscopy: SEM, freeze fracture</li><li>5. Electron microscopy: TEM</li><li>6. Fluorescence microscopy</li><li>7. Confocal microscopy: imaging live cells.</li><li>8. FRET</li><li>9. FRAP</li><li>10. Flow cytometry and Fluorescence assisted cell sorting (FACS)</li><li>11. Cell counting -hemocytometer</li><li>12. Plant tissue culture.</li><li>13. Animal tissue culture</li><li>14. Insect tissue culture</li><li>15. Methods of cell disruption and fractionation and isolation of organelles</li></ol>



<b>PRACTICAL (Each practical topic consists of 3 Experiments, Total 15 Expts)</b>	
<b>CREDIT 4</b>	<b>Quantitation of Biomolecules by Spectroscopy (30 h)</b>
	<ol style="list-style-type: none"> <li>1 Absorption spectrum of tyrosine, determination of molar extinction coefficient,</li> <li>2 Estimation of concentration of biomolecules based on Beer Lambert's Law</li> <li>3 Estimation of inorganic phosphate by Fiske-Subbarow method</li> <li>4 Estimation of protein by Biuret method</li> <li>5 Estimation of protein by Lowry method</li> </ol>
<b>CREDIT 5</b>	<b>Separation of Biomolecules by Chromatography (30 h)</b>
	<ol style="list-style-type: none"> <li>1 Separation of plant pigments by paper chromatography</li> <li>2 1-D and 2-D paper chromatography of amino acids</li> <li>3 TLC of plant pigments and lipids</li> <li>4 Anion/ Cation-exchange capacity of resin</li> <li>5 Separation of amino acids by ion-exchange chromatography</li> </ol>

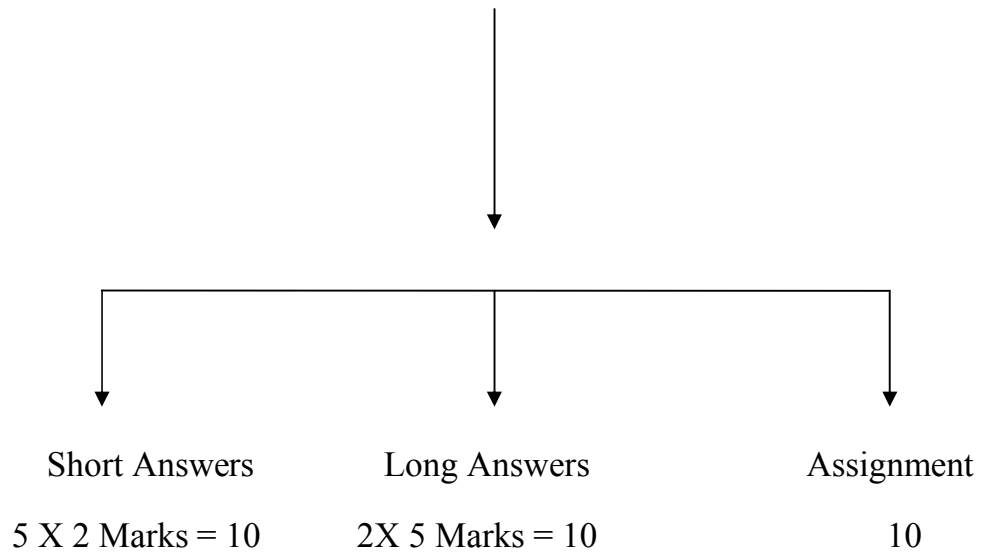
## **SCHEME OF EVALUATION**

Each paper - Theory = 100 marks, Practical = 50 Marks.

Theory: Internal assessment=30 Marks + Semester end examination = 70 Marks

### **THEORY**

**INTERNAL ASSESSMENT= 30 Marks**



**SEMESTER END EXAMINATION =70 Marks**

**70 Marks -**

**3 UNITS**

**PART – A**

Answer all questions

5 x 5 = 25 Marks

1.

2.

3.

4.

5.

**PART- B**

Answer All Questions

3 X 15=45 Marks

6 A

or

B

7 A

or

B

8 A

or

B